Francisella tularensis is a bacterium that induces the zoonotic disease tularemia. In the course of infection, F. tularensis bacteria invade erythrocytes, a phenomenon that heightens the colonization of ticks after a blood meal. To better understand the mechanism of erythrocyte invasion, we hypothesized that transcription of bacterial genes significant in erythrocyte invasion would be upregulated upon exposure to these host cells. An RNA-seq unveiled that transcription of 7% of F. tularensis genes augment when in erythrocyte presence. Of these, we pinpointed a putative transcriptional regulator, FTL_1199. The goal was to delete FTL_1199 in F. tularensis LVS. SOE PCR amplified and duplicated the up and downstream regions of the target gene in tandem into a shuttle vector that is insecure within F. tularensis. This newly generated plasmid, pDEL1199, was mobilized inside of F. tularensis by conjugation. Merodiploid strains generated by homologous recombination were isolated and transformed with pGUTS. Expression of I-Sce1 within the merodiploid produces a double-stranded break. This breakage resulted in a second recombination that either ensued to wild-type or deletion of FTL_1199 deduced through a PCR. Finally, in ΔFTL_1199 strains, pGUTS was cured by successive cultivation in the absence of selection followed by replica-plating on chocolate II agar ± kanamycin. Gentamicin protection assays showed reduced levels of erythrocyte invasion for F. tularensis ΔFTL_1199 compared to wild type bacteria. However, complementation of FTL_1199 to the deletion mutant restored this strain’s ability to invade red blood cells. These findings demonstrate that FTL_1199 is important for erythrocyte invasion by F. tularensis.

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